

Mouse Anti-KLH IgG ELISA

Life Diagnostics, Inc., Catalog Number: KLHG-1

Mouse Anti-KLH IgG ELISA

INTRODUCTION

Measurement of KLH (keyhole limpet hemocyanin) induced anti-KLH antibody levels allows quantitative evaluation of the immune response (ref 1). This ELISA is designed for the rapid and quantitative measurement of mouse anti-KLH IgG levels in serum or plasma. A companion ELISA, catalog number KLHM-1, is designed for measurement of mouse anti-KLH IgM levels.

PRINCIPLE OF THE ASSAY

The mouse anti-KLH IgG test kit is based on a solid phase enzyme-linked immunosorbent assay (ELISA). The assay uses KLH for solid phase (microtiter wells) immobilization and horseradish peroxidase (HRP) conjugated anti-mouse IgG antibodies for detection. Test serum or plasma samples are diluted and incubated in the microtiter wells for 1 hour. The microtiter wells are subsequently washed, and HRP conjugate is added and incubated for 45 minutes. Anti-KLH IgG molecules are thus sandwiched between immobilized KLH and the detection antibody conjugate. The wells are then washed to remove unbound HRP-labeled antibodies, and TMB Reagent is added and incubated for 20 minutes at room temperature. This results in the development of a blue color. Color development is stopped by the addition of Stop Solution, changing the color to yellow. Optical density is measured spectrophotometrically at 450 nm. The concentration of anti-KLH IgG is proportional to the optical density of the test sample.

MATERIALS AND COMPONENTS

Materials provided with the kit:

- KLH Coated 96-well Plate (provided as 12 strips of 8 wells)
- Enzyme Conjugate Reagent, 11 ml
- Reference Standard¹ (lyophilized)
- 20x Wash Solution: TBS50-20, 50 ml
- Diluent: YD60-1, 60 ml
- TMB Reagent (One-Step): TMB11-1, 11 ml
- Stop Solution (1N HCl): SS11-1, 11 ml

Materials required but not provided:

- Precision pipettes and tips
- Distilled or deionized water
- Polypropylene or glass tubes
- Vortex mixer
- Absorbent paper or paper towels
- Micro-plate incubator/shaker mixing speed of 150 rpm
- Plate washer
- Plate reader with an optical density range of 0-4 at 450 nm
- Graph paper (PC graphing software is optional)

STORAGE

- The reference standard should be stored at -20°C for optimal stability
 - All remaining kit components should be stored at 4°C
- The microtiter plate should be kept in a sealed bag with desiccant to minimize exposure to damp air. The test kit will remain stable for six months from the date of purchase provided that the components are stored as described above.

GENERAL INSTRUCTIONS

1. Please read and understand the instructions thoroughly before using the kit.
2. This kit is intended for measurement of anti-KLH IgG levels in mouse serum or plasma obtained 14-days after immunization with KLH.
3. All reagents should be allowed to reach room temperature (25°C) before use.
4. The optimal sample dilution should be determined empirically. However, studies performed at Life Diagnostics, Inc. indicate that an initial sample dilution of 20,000-fold works well for most 14-day post-immunization samples. Please do not use dilutions less than 25-fold.
5. Optimal results are achieved if, at each step, reagents are pipetted into the wells of the microtiter plate within 5 minutes.

WASH SOLUTION PREPARATION

The wash solution is provided as a 20x stock. Prior to use dilute the contents of the bottle (50 ml) with 950 ml of distilled or deionized water.

STANDARD PREPARATION

1. The mouse anti-KLH IgG standard is provided as a lyophilized stock. Reconstitute with 100 µl of distilled or deionized water (**the reconstituted standard is stable at 4°C for one week but should be aliquoted and frozen at -20°C after reconstitution if future use is intended**).
2. Label 5 polypropylene or glass tubes as 100, 50, 25, 12.5 and 6.25 units/ml (u/ml).
3. Into the tube labeled 100 u/ml, pipette the volume of diluent detailed on the anti-KLH IgG standard vial label. Then add the indicated volume of anti-KLH IgG standard (shown on the anti-KLH IgG standard vial label) and mix gently. This provides the 100 u/ml standard.
4. Dispense 250 µl of diluent into the tubes labeled 50, 25, 12.5, and 6.25 u/ml.
5. Prepare a 50 u/ml standard by diluting and mixing 250 µl of the 100 u/ml standard with 250 µl of diluent in the tube labeled 50 u/ml.
6. Similarly prepare the 25, 12.5, and 6.25 u/ml standards by serial dilution.

¹ Mouse anti-KLH IgG levels are measured in nominal units and are calibrated with reference anti-KLH mouse serum at Life Diagnostics, Inc.

SAMPLE PREPARATION

General Note: Studies at Life Diagnostics, Inc. indicate that anti-KLH IgG is present in serum from KLH immunized mice at concentrations of approximately 750,000 u/ml. To obtain values within range of the standard curve, we suggest that samples initially be diluted 20,000-fold using the following procedure for each sample to be tested:

1. Dispense 248 μ l and 318 μ l of diluent into separate tubes.
2. Pipette and mix 2 μ l of the serum sample into the tube containing 248 μ l of diluent. This provides a 125-fold diluted sample.
3. Mix 2 μ l of the 125-fold diluted sample with the 318 μ l of diluent in the second tube. This provides a 20,000-fold dilution of the sample.
4. Repeat this procedure for each sample to be tested.

ASSAY PROCEDURE

1. Secure the desired number of coated wells in the holder.
2. Dispense 100 μ l of standards and diluted samples into the wells (we recommend that samples be tested in duplicate).
3. Incubate on an orbital micro-plate shaker at 100-150 rpm at room temperature (25°C) for 1 hour.
4. Aspirate the contents of the microtiter wells and wash the wells 5 times with 1x wash solution using a plate washer (400 μ l/well). The entire wash procedure should be performed as quickly as possible.
5. Strike the wells sharply onto absorbent paper or paper towels to remove all residual wash buffer.
6. Add 100 μ l of enzyme conjugate reagent into each well.
7. Incubate on an orbital micro-plate shaker at 100-150 rpm at room temperature (25°C) for 45 minutes.
8. Wash as detailed in 4 to 5 above.
9. Dispense 100 μ l of TMB Reagent into each well.
10. Gently mix on an orbital micro-plate shaker at 100-150 rpm at room temperature (25°C) for 20 minutes.
11. Stop the reaction by adding 100 μ l of Stop Solution to each well.
12. Gently mix. *It is important to make sure that all the blue color changes to yellow.*
13. Read the optical density at 450 nm with a microtiter plate reader within 5 minutes.

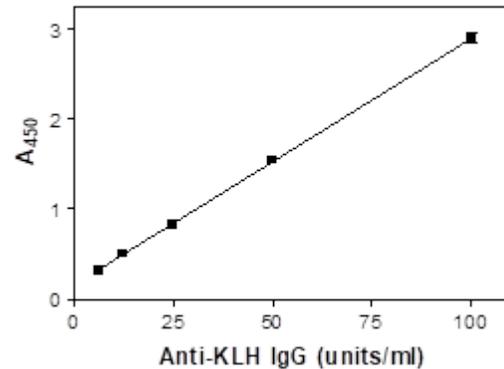
CALCULATION OF RESULTS

1. Calculate the average absorbance values (A_{450}) for each set of reference standards and samples.
2. Construct a standard curve by plotting the mean absorbance obtained from each reference standard against its concentration in ng/ml on linear graph paper, with absorbance values on the vertical or Y-axis and concentrations on the horizontal or X-axis.
3. Using the mean absorbance value for each sample, determine the corresponding concentration of anti-KLH IgG in u/ml from the standard curve.
4. Multiply the derived concentration by the dilution factor to determine the actual concentration of anti-KLH IgG in the serum/plasma sample.
5. PC graphing software may be used for the above steps.
6. If the OD_{450} values of samples fall outside the standard curve when tested at a 20,000-fold dilution, samples should be diluted appropriately and re-tested.

TYPICAL STANDARD CURVE

A typical standard curve with optical density readings at 450nm on the Y-axis against anti-KLH IgG concentrations on the X-axis is shown below. This curve is for the purpose of illustration only and should not be used to calculate unknowns.

Anti-KLH IgG (u/ml)	A_{450}
100	2.903
50	1.542
25	0.821
12.5	0.509
6.25	0.312



LIMITATIONS OF THE PROCEDURE

1. Reliable and reproducible results will be obtained when the assay procedure is carried out with a complete understanding of and in accordance with the instructions detailed above.
2. The wash procedure is critical. Insufficient washing will result in poor precision and falsely elevated absorbance readings.

REFERENCES

1. Wada H, Noguchi Y, Marino MW, Dunn AR and Old LJ. T-cell functions in granulocyte/macrophage colony-stimulating factor deficient mice. Proc Natl Acad Sci. 94:12557-61 (1997)

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