

Monkey IgA ELISA

Life Diagnostics, Inc., Catalog Number: IGA-3

Monkey IgA ELISA

INTRODUCTION

The monkey IgA ELISA kit is designed for measurement of IgA in old world monkey serum or plasma. The assay uses goat anti-monkey IgA for solid phase (microtiter wells) immobilization and horseradish peroxidase (HRP) conjugated goat anti-monkey IgA antibodies for detection. Both capture and detection antibodies were cross absorbed on monkey IgG and IgM agarose columns, thereby ensuring specificity for IgA. Cross-reactivity with immunoglobulins from other species has not been investigated.

IgA is present in monkey serum, milk, and mucosal secretions at concentrations up to 2 mg/ml.^{1,2}

PRINCIPLE OF THE ASSAY

Test samples are diluted and incubated in the microtiter wells for 45 minutes alongside monkey IgA standards. The microtiter wells are subsequently washed and HRP conjugate is added and incubated for 45 minutes. IgA molecules are sandwiched between the immobilization and detection antibodies. The wells are then washed to remove unbound HRP-labeled antibodies. TMB Reagent is added and incubated for 20 minutes at room temperature. This results in the development of a blue color. Color development is stopped by the addition of Stop Solution, changing the color to yellow. Optical density is measured spectrophotometrically at 450 nm. The concentration of IgA is proportional to the optical density of the test sample and is derived from a standard curve.

MATERIALS AND COMPONENTS

Materials provided with the kit:

- Anti monkey IgA coated 96-well plate (12 strips of 8 wells)
- HRP Conjugate Reagent, 11 ml
- Reference standard (lyophilized)
- 20x Wash Solution: TBS50-20, 50 ml
- 10x Immunoglobulin Diluent: RD25-10, 25 ml
- TMB Reagent (One-Step): TMB11-1, 11 ml
- Stop Solution (1N HCl): SS11-1, 11 ml

Materials required but not provided:

- Precision pipettes and tips
- Distilled or deionized water
- Polypropylene or glass tubes
- Vortex mixer
- Absorbent paper or paper towels
- Micro-Plate incubator/shaker mixing speed of 150 rpm
- Plate washer
- Plate reader with an optical density range of 0-4 at 450nm
- PC graphing software

STORAGE OF THE TEST KIT

The test kit will remain stable for six months from the date of purchase provided that the components are stored at 4°C. The microtiter plate should be kept in a sealed bag with desiccant to minimize exposure to damp air.

GENERAL INSTRUCTIONS

1. Please read and understand the instructions thoroughly before using the kit.
2. All reagents should be allowed to reach room temperature (25°C) before use.
3. Optimal results are achieved if, at each step, reagents are pipetted into the wells of the microtiter plate within 5 minutes.

DILUENT PREPARATION

The diluent is provided as a 10x stock. Prior to use, estimate the final volume of diluent required for your assay and dilute one (1) volume of the 10x stock with nine (9) volumes of distilled or deionized water.

WASH SOLUTION PREPARATION

The wash solution is provided as a 20x stock. Prior to use, dilute the contents of the bottle (50 ml) with 950 ml of distilled or deionized water.

STANDARD PREPARATION

1. The IgA standard is provided as a lyophilized stock. Reconstitute with 1.0 ml of distilled or deionized water (***the reconstituted standard is stable at 4°C for one week but should be aliquoted and frozen at -20°C after reconstitution if future use is intended***).
2. Label 6 polypropylene or glass tubes as 125, 62.5, 31.25, 15.63, 7.81 and 3.91 ng/ml.
3. Into the tube labeled 125 ng/ml, pipette the volume of diluent detailed on the IgA standard vial label. Then add the indicated volume of IgA standard (shown on the IgA standard vial label) and mix gently. This provides the 125 ng/ml standard.
4. Dispense 250 µl of diluent into the tubes labeled 62.5, 31.25, 15.63, 7.81 and 3.91 ng/ml.
5. Prepare a 62.5 ng/ml standard by diluting and mixing 250 µl of the 125 ng/ml standard with 250 µl of diluent in the tube labeled 62.5 ng/ml.
6. Similarly prepare the 62.5, 31.25, 15.63, 7.81 and 3.91 ng/ml standards by serial dilution.

SAMPLE PREPARATION

General Note: IgA is typically present in monkey serum or plasma at concentrations of approximately 2 mg/ml. To obtain values within range of the standard curve, we suggest that samples initially be diluted 50,000-fold using the following procedure for each sample to be tested:

1. Dispense 498 µl and 497.5 µl of 1x diluent into separate tubes.
2. Pipette and mix 2 µl of the serum/plasma sample into the tube containing 498 µl of diluent. This provides a 250-fold diluted sample.
3. Mix 2.5 µl of the 250-fold diluted sample with the 497.5 µl of diluent in the second tube. This provides a 50,000-fold dilution of the sample.
4. Repeat this procedure for each sample to be tested.

Body fluids other than serum or plasma will likely have lower IgA levels than those found in serum. Optimal dilutions of such samples should be determined empirically.

ASSAY PROCEDURE

1. Secure the desired number of coated wells in the holder.
2. Dispense 100 μ l of standards and diluted samples into the wells (we recommend that samples be tested in duplicate).
3. Incubate on an orbital micro-plate shaker at 100-150 rpm at room temperature (25°C) for 45 minutes.
4. Aspirate the contents of the microtiter wells and wash the wells 5 times with 1x wash solution using a plate washer (400 μ l/well). The entire wash procedure should be performed as quickly as possible.
5. Strike the wells sharply onto absorbent paper or paper towels to remove all residual wash buffer.
6. Add 100 μ l of enzyme conjugate reagent into each well.
7. Incubate on an orbital micro-plate shaker at 100-150 rpm at room temperature (25°C) for 45 minutes.
8. Wash as detailed in 4 to 5 above.
9. Dispense 100 μ l of TMB Reagent into each well.
10. Gently mix on an orbital micro-plate shaker at 100-150 rpm at room temperature (25°C) for 20 minutes.
11. Stop the reaction by adding 100 μ l of Stop Solution to each well.
12. Gently mix. *It is important to make sure that all the blue color changes to yellow.*
13. Read the optical density at 450 nm with a microtiter plate reader within 5 minutes.

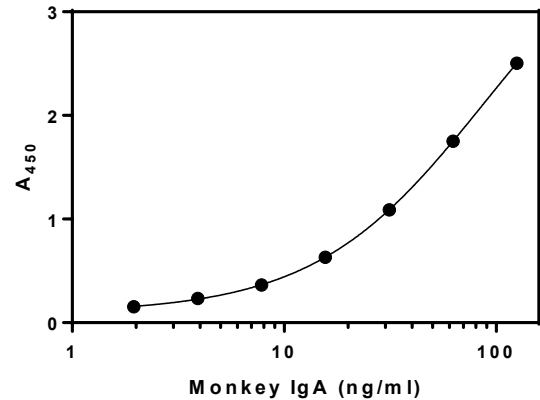
CALCULATION OF RESULTS

1. Using graphing software, construct a standard curve by plotting the absorbance values for the standards versus the \log_{10} of the IgA concentration.
2. Derive the corresponding concentration of IgA in the samples from the standard curve (remember to derive the concentration from the antilog).
3. Multiply the derived concentration by the dilution factor to determine the actual concentration of IgA in the sample.
4. If the absorbance values of diluted samples fall outside the standard curve, samples should be diluted appropriately and re-tested.

TYPICAL STANDARD CURVE

A typical standard curve with optical density readings at 450nm on the Y-axis against \log_{10} IgA concentrations on the X-axis is shown below. This curve is for the purpose of illustration only and should not be used to calculate unknowns. Each user should obtain his or her data and standard curve in each experiment.

IgA (ng/ml)	A_{450}
125	2.505
62.5	1.752
31.25	1.090
15.63	0.632
7.81	0.364
3.91	0.234



LIMITATIONS OF THE PROCEDURE

1. Reliable and reproducible results will be obtained when the assay procedure is carried out with a complete understanding of and in accordance with the instructions detailed above.
2. The wash procedure is critical. Insufficient washing will result in poor precision and falsely elevated absorbance readings.

REFERENCES

1. MF Cole & WH Bowen. Immunoglobulins A, G, and M in serum and in some secretions of monkeys (*Macaca fascicularis* syn. *irus*). *Infection and Immunity* 13:1354-1359 (1976)
2. F Xusheng Lu, et al. Immunoglobulin concentrations and antigen-specific antibody levels in cervicovaginal lavages of rhesus macaques are influenced by the stage of the menstrual cycle. *Infection and Immunity* 67:6321-6328 (1999)

Rev 10202020

For technical assistance please email us at
techsupport@lifediagnosics.com